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2	Article Sub-Title		
3	Article Copyright - Year	Springer-Verlag Wien 2013 (This will be the copyright line in the final PDF)	
4	Journal Name	Protoplasma	
5		Family Name	Nick
6		Particle	
7		Given Name	Peter
8	Corresponding	Suffix	
9	Author	Organization	
10		Division	
11		Address	Karlsruhe, Germany
12		e-mail	peter.nick@bio.uka.de
13		Received	
14	Schedule	Revised	
15		Accepted	
16	Abstract		
17	Keywords separated by ' - '		
18	Foot note		

information

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Protoplasma DOI 10.1007/s00709-013-0551-6

EDITORIAL

## A cell biologist on Mars—the exotic world of algal cells

Peter Nick

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In his famous book "An anthropologist on Mars," Olivers

Sacks presents exotic cases of neural disorders, and while

describing the seemingly extraterrestrial world of these patients,

he unveils the hidden wonders in the brain of the "normal"

majority. In the history of science, very often, the fascination for

the abnormal, exotic and mysterious has paved the way for a

deeper understanding of the apparently "conventional." Cell

biology is no exception to this rule. It seems that the era of

channelling towards a few model systems accessible for mo-

lecular approaches is now progressively replaced by a wider

approach towards the variations of life. The renewed interest for

"off-track" models is certainly fuelled by the advances in se-

quencing technologies that allow molecular insights even for

models carried by only smaller research communities. Why are

these "off-track" models important for science? New complex

traits, such as new morphogenetical programmes, physiological

achievements or metabolic pathways seem to emerge mainly

from functional shifts of the underlying cellular and molecular

events. These shifts would not be possible in systems where a

function is exactly fitted to its current function. However, to

quote a famous metaphor by François Jacob (1977), evolution

is tinkering (and therefore neither design nor intelligent...),

which means that most proteins are not exactly fitted, but in

addition to their main function, maintain the ability for other

hidden functions. In a different functional context, these moon-

lighting functions (Kurakin 2005) can take the lead. The heter-

ogenous group of "algae" comprising highly diverse life forms

with a mostly phototropic lifestyle provides good examples to

illustrate the case, and the study of these seemingly extraterres-

trial models might help us to get track of one or the other

"moonlighting function" hidden in the cell biology of "higher"

life forms. The current issue assembles several fine examples of

generation of new life forms through endocytosis. The review by

Stork et al. (2013) in the current issue gives a survey on the

The tinkering approach of evolution is neatly illustrated by the

current state of the highly dynamic and highly controversial field 47 of secondary endosymbiosis with green or red algae (that unlike 48stated in the last editorial to the current state of knowledge arise 49from primary endosymbiosis, as stated by attentive readers). 50Secondary endosymbiosis gave birth to many important groups 51of "algae" including heterokontophytes (for recent review, see 52Beakes et al. 2011), haptophytes, cryptophytes, and flagellate 53parasites. A core element for the functional integration of for-54merly independent organisms into a novel supraorganism is the 55protein transport across the membranes between the symbiotic 56partners. Especially the transport across the second outermost 57plastid membrane that is derived from the former endosymbiont 58represents a major challenge. The review shows how the ER-59dependent degradation machinery of the endosymbiont was 60 remodelled and relocated to generate a novel transport machinery 61 carrying the new task of importing proteins across the former cell 62 membrane. Thus, evolution is not only "tinkering" by recycling 63 pre-existing machineries into a new functional context, evolution 64 apparently is also playing with modular "LEGO" bricks of 65 function. The challenge will be to understand how tinkering is 66 reflected in the networks of gene regulation. 67

One of the most mysterious and fascinating groups of "algae" 68 are the diatoms that are treated by even two research publications 69 in the current issue. Already by their lifestyle as pure diplonts, 70they appear a bit alien within the plant kingdom and their 71mysterious mode of locomotion, as well as their beautiful silica 72shells (that have inspired art and architecture alike) support the 73impression of a highly exotic form of life. Although minute in 74size, diatoms are of global impact and account for an estimated 75one fourth of the global cycles for silicium and carbon. Despite 76 this impact, the question, how the unique, filigrane and species-77 specific silica patterns (the so called *frustulae*) are actually 78formed, has remained enigmatic. It was in this journal, where 79 for the first time the electron microscopical detection of minute 80 particles in the cytoplasm of diatoms was reported and these 81 particles were proposed to represent transport vesicles for silicon 82 (Schmid and Schulz 1979). This observation is now revisited by 83 the work of Annenkov et al. (2013) in the current issue, but using 84 a fluorescent dye specific for growing siliceous frustulae such 85 that the function of these vesicles could be addressed by life-cell 86

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algal cell biology:

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imaging. They used silicon starvation to achieve a synchronisa-87 tion of silicon deposition-a simple, but smart approach from 88 classical physiology to resolve the temporal sequence of causal 89 90 chains. They can show that within minutes after readdition of 91silicon together with the specific dye, fluorescent vesicles appear in the cytoplasm and subsequently disappear again while sili-92ceous valves are deposited. This study not only supports the 93 classical hypothesis by Schmid and Schulz (1979) that the cyto-94plasmic particles are silicon transport vesicles but also resolves a 95two-stage mechanism of rapid Si uptake followed by a slower 96 deposition of silicon into the growing frustulae. A second mys-97 98 tery of diatoms, their enigmatic mode of locomotion, is addressed by the work of Wang et al. (2013) in the current issue. Again, the 99starting point was an observation published in this journal (Edgar 1001983): diatoms moving over benthic surfaces secrete consider-101 able volumes of mucilage, and the so called Edgar model as-102103sumed that motility is generated by a conformational change of the adhesive mucilage during its secretion. The direction of 104105secretion was thought to be confined by actin to one end of the cell such that it is pushed forward. This model, although 106appearing plausible, suffers from several weak points that stim-107 ulated the authors to search for a better model. For instance, it is 108 109 not clear how sign reversals of movement might be generated by the Edgar mechanism. The authors analysed cell structure, move-110ment, mucilage and bending deformation for the benthic pennate 111 112diatom Navicula and come up with a new model, which is also supported by biophysical modelling, where pseudopodia pro-113trude from the frustulae attach to the surface through secreted 114 115extracellular polymers and where other pseudopodia push 116 against the substrate providing the driving force for locomotion.

Brown algae harbour the most differentiated life forms among 117118 the algae and can produce highly sophisticated architectures. Due to this complexity, these architectures have to be tuned with 119respect to their environment mainly with the distribution of light 120 121in their benthic habitat. Therefore, the zygotes and spores of brown algae are photosensitive and adjust axis and polarity with 122123the direction of light during a photosensitive period. The 124photopolarisation of the Fucus zygote has been extensively studied in the second half of the last century and led to a model, 125where light-triggered calcium currents drive a transcellular gra-126dient of actin-dependent calcium channels such that initial weak 127gradients are self-amplified into a robust polarity by a process 128that had been termed "self-electrophoresis." A drawback of 129130Fucus system has been the lack of molecular information and during the abovementioned channelling upon few models; this 131132beautiful system for cellular development has been mostly abandoned. It seems that the new move towards genomics of "off-133track" models will also bring a renaissance of brown algae as 134systems for plant development. The related species Ectocarpus 135siliculosus has been sequenced and is currently pushed as new 136137genetic model. Here, the mitospores can be polarised by light in a manner that resembles the situation in the Fucus zygote. The 138work by Green et al. (2013) in the current issue investigates the 139195

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role of microtubules in polarisation. They show that pharmaco-140logical manipulation of microtubules impairs polarisation, an 141 even stronger inhibition can be produced by inhibitors of vesicle 142flow. By confocal microscopy, they visualise a radial array of 143microtubules emanating from an organelle that resembles a 144 centrosome and is localised close to the nuclear rim. Upon 145germination, this array gives rise to longitudinal microtubules 146 that reach into the bulging filament. Within the filament, mitotic 147 spindles are aligned with the axis of growth. The characterisation 148 of these cellular events is important as phenomenological frame-149work to interpret phenotypes for future studies on the functional 150genetics of polarity induction. Ectocarpus differs in a curious 151detail from the classical Fucus system: polarity induction in 152Fucus is a matter of actin and microtubules seem to be 153dispensible in this context. In the Ectocarpus system, it is 154microtubules that take the lead-the authors suggest that this 155difference might be linked with the presence of a centrosome in 156case of the Ectocarpus mitospores, whereas in Fucus, the cen-157trosomes are paternally inherited (Motomura 1994). This detail 158shows that even in cell biology, often perceived as science 159searching for general laws, details and particularities of models 160 are relevant and can when they are seriously considered by 161 comparison allow for new mechanistic insights (a topic 162addressed in a previous editorial, Nick 2009). 163

Conflict of interest The author declares that there is no conflict of interest. 165

#### References

- Annenkov V. Basharina TN. Danilovtseva EN. Grachev MA (2013) Putative 168silicon transport vesicles in the cytoplasm of the diatom Synedra acus 169during surge uptake of silicon. Protoplasma, current issue 170Beakes GW, Glockling SL, Sekimoto S (2011) The evolutionary phylog-171
- eny of the oomycete "fungi". Protoplasma 249:3-19 172173
- Edgar L (1983) Mucilage secretions of moving diatoms. Protoplasma 118:44-48
- Green JJ, Cordero DF, Peters NT, Logan KO, Kropf DL (2013) Dynamic 175Microtubules and Endomembrane Cycling Contribute to Polarity 176Establishment and Early Development of Ectocarpus Mitospores. 177Protoplasma, current issue 178179
- Jacob F (1977) Evolution and Tinkering. Science 196:1161-1166
- Kurakin A (2005) Self-organisation vs. Watchmaker: stochastic dynamics of cellular organisation. Biol Chem 386:247-254
- Motomura T (1994) Electron and immunofluorescence microscopy of the 182fertilization of Fucus distichus (Fucales, Phaeophyceae). Protoplasma 183178:97-110 184185
- Nick P (2009) Comparing is worth the effort-lessons from mitosis. Protoplasma 237:1-2
- Schmid A-MM, Schulz D (1979) Wall morphogenesis in diatoms: depo-187 sition of silica by cytoplasmic vesicles. Protoplasma 100:267-288 188
- Stork S, Lau J, Moog D, Maier U (2013) Three old and one new: Protein 189import into red algal-derived plastids surrounded by four mem-190191 branes. Protoplasma, current issue
- Wang JD, Cao Sh, Du Ch, Chen DR (2013) Underwater Locomotion 192Strategy by a Benthic Pennate Diatom Navicula sp. Protoplasma, 193current issue 194

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